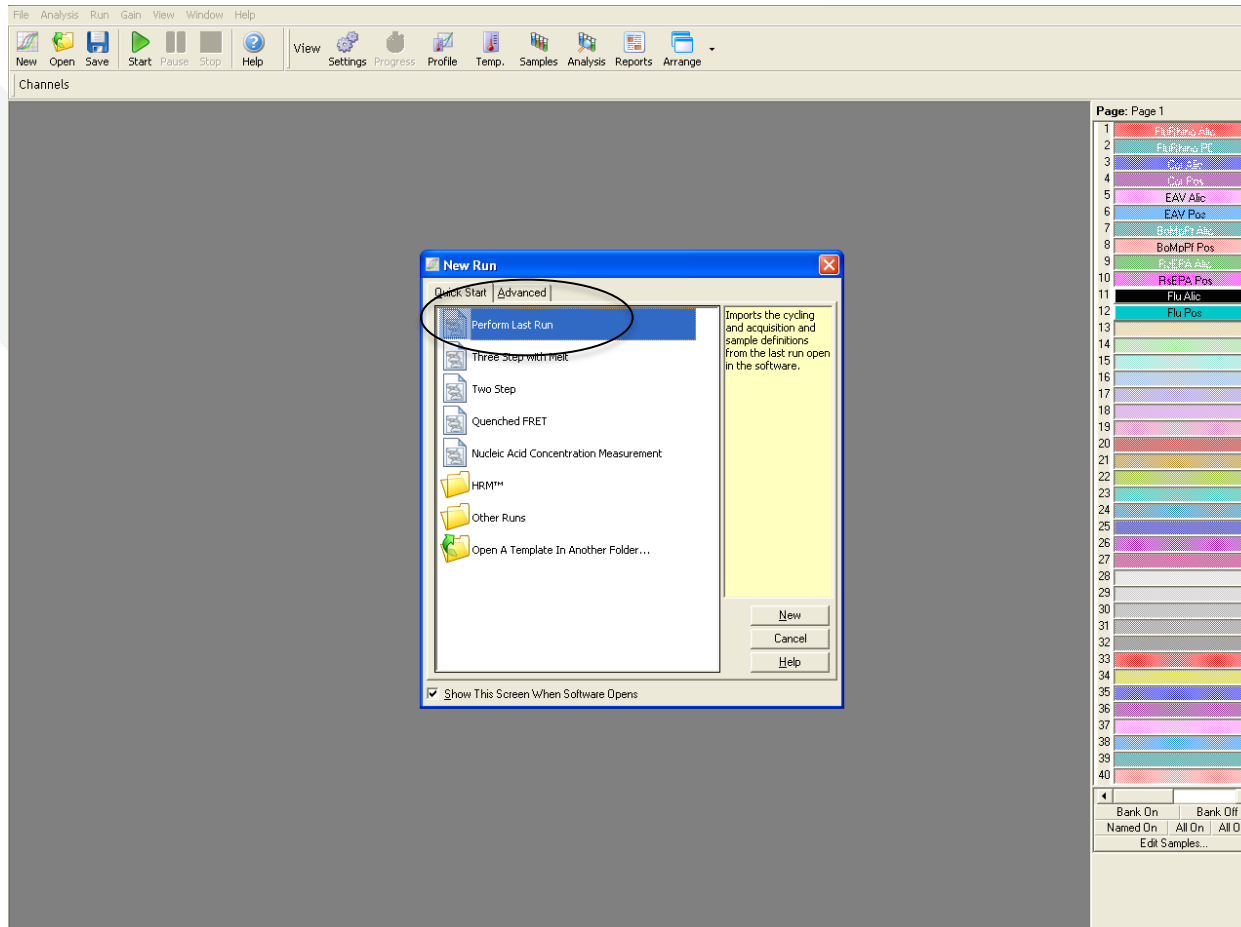
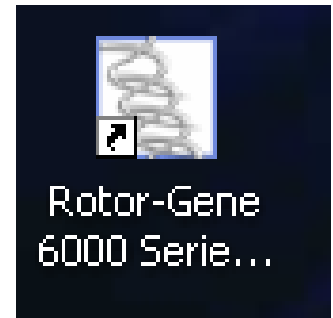


Set-up, programming and analysis: RotorGene 3000/6000, Rotor-Gene Q

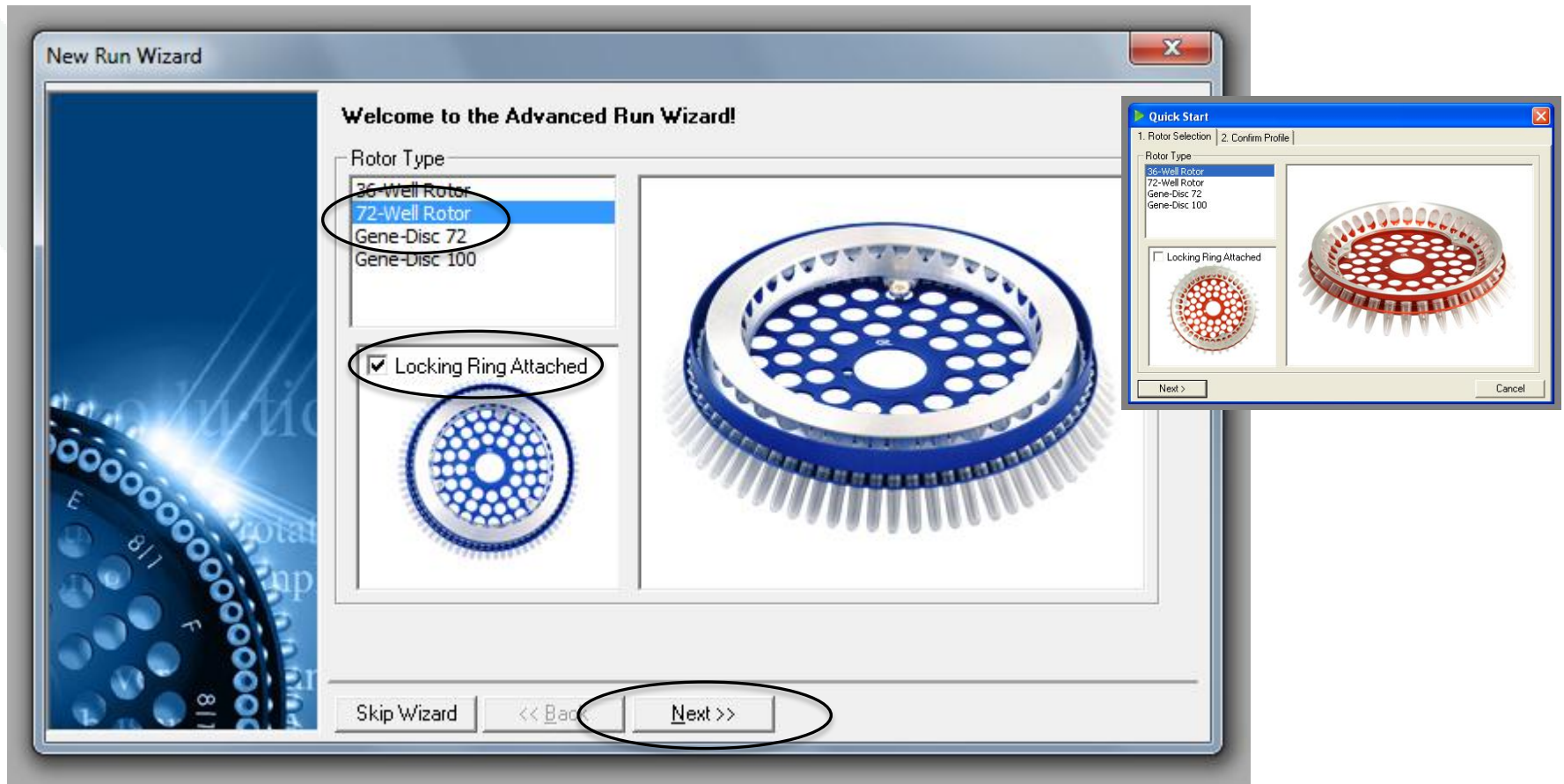


How to start a RotorGene run.



1. Start the Rotor-Gene software.
2. Choose: Advanced/ Perform Last Run and press New.

How to choose the rotor.



1. Choose the Rotor you use (36 or 72 tubes) in the Rotor Selection field.
2. Tick Locking Ring Attached and press Next.

How to edit the set up of a run method.

New Run Wizard

This screen displays miscellaneous options for the run. Complete the fields, clicking Next when you are ready to move to the next page.

Operator : nou

Notes :

Reaction Volume (µL): 25

Sample Layout : 1, 2, 3...

Enter any additional notes for the run in this text box.

Skip Wizard << Back Next >>

1. You can name the operator and write down some notes.
2. Choose 25ul as Reaction Volume and define your Sample Layout (1,2,3...)
3. Press Next.

How to edit the set up of a run method.

Temperature Profile :

Channel Setup :

Name	Source	Detector	Gain
Green	470nm	510nm	5
Yellow	530nm	555nm	5
Orange	585nm	610nm	5
Red	625nm	660nm	5
Crimson	680nm	710hp	7

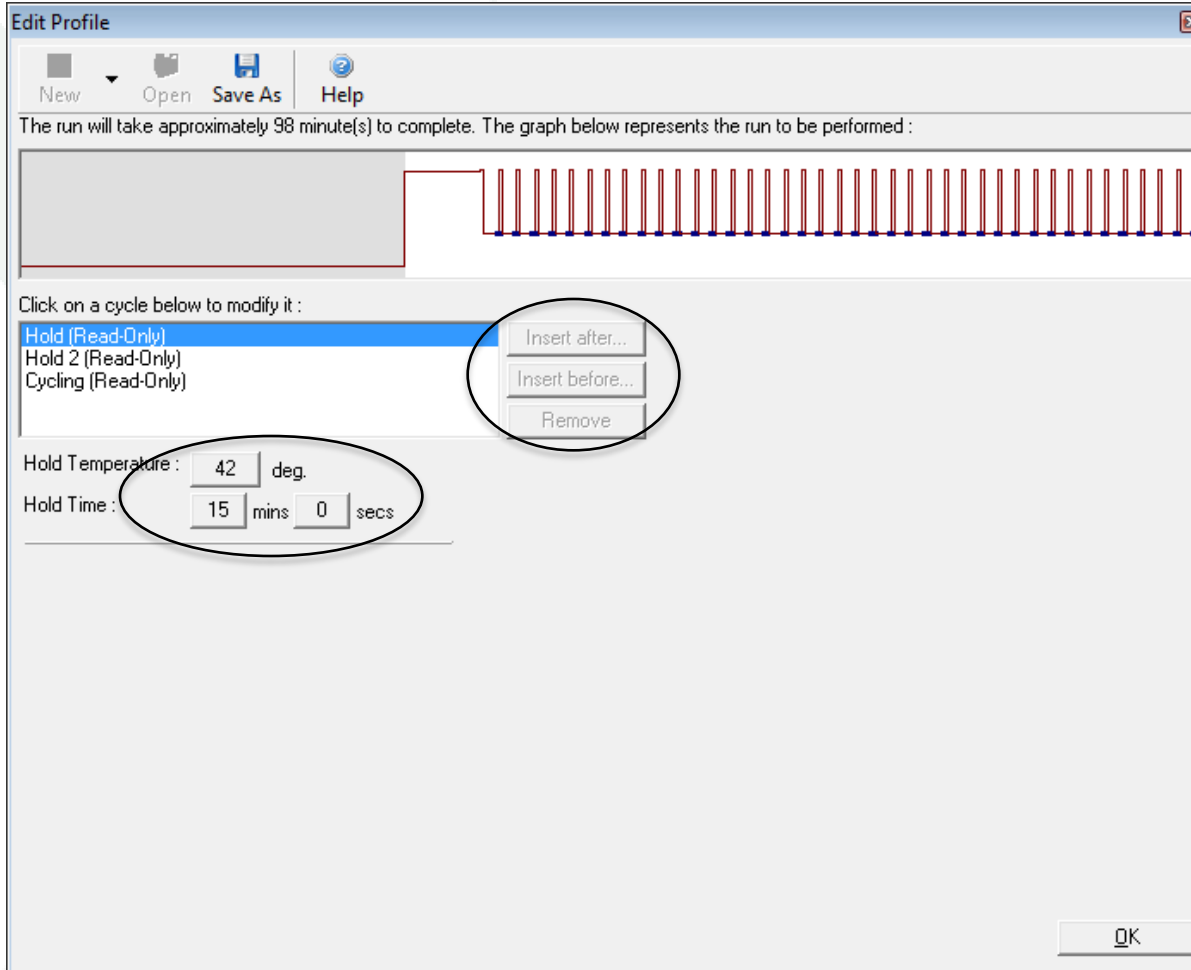
Gain Optimisation...

Gain for Green

5

1. Tick the Green Channel.
2. Press Edit Gain and the small window Gain for Green appears.
3. Set the gain on 5.
4. Repeat the same for Yellow, Orange and Red.
5. Press Edit Profil.

How to edit the set up of a run method.



Confirm profile:

Hold 42°C -> 15min

Hold2 94°C -> 3min

Cycling 94°C -> 8sec

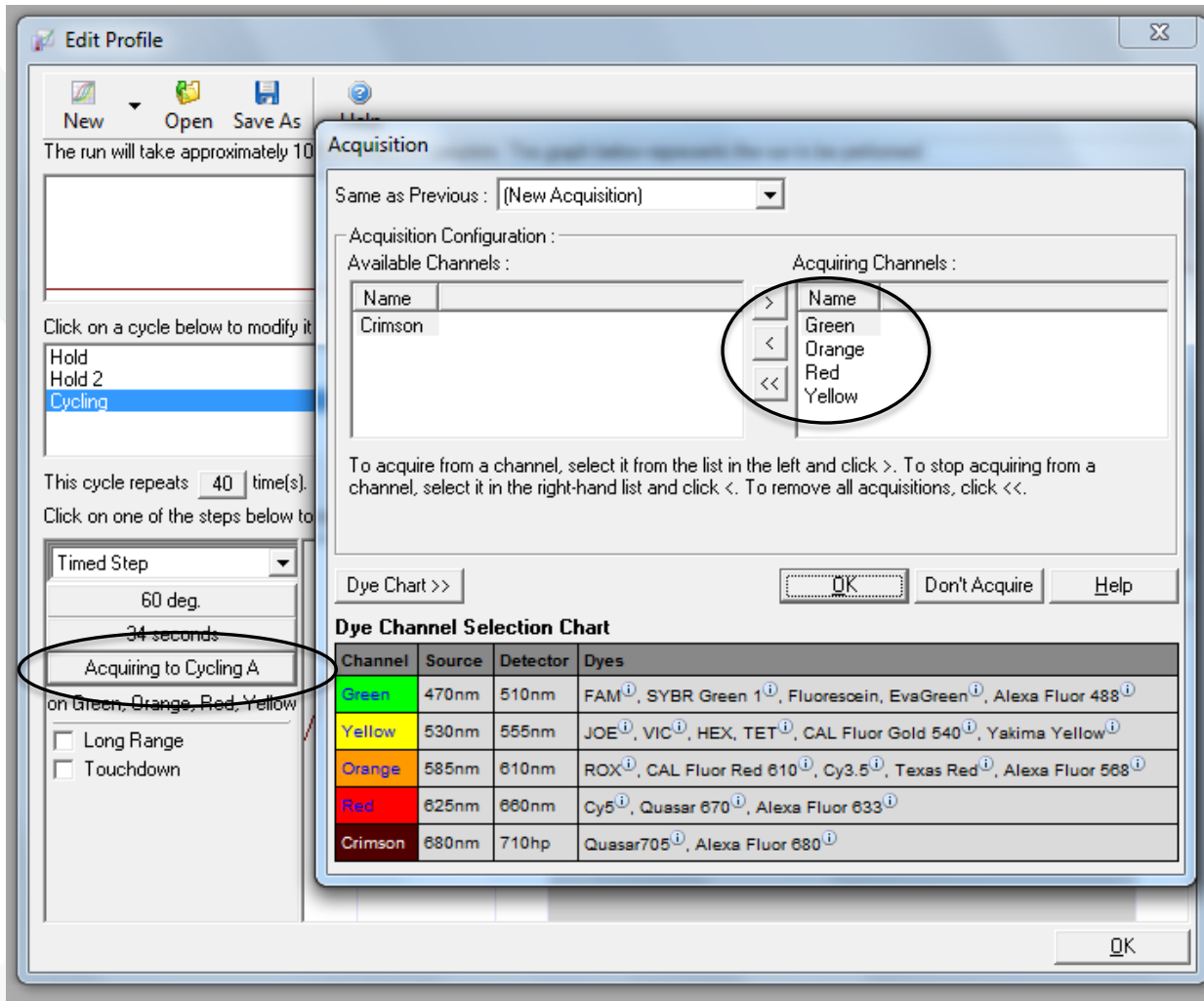
60°C -> 34sec

Aquiring to Cycle A

Cycle repeat: 40 times

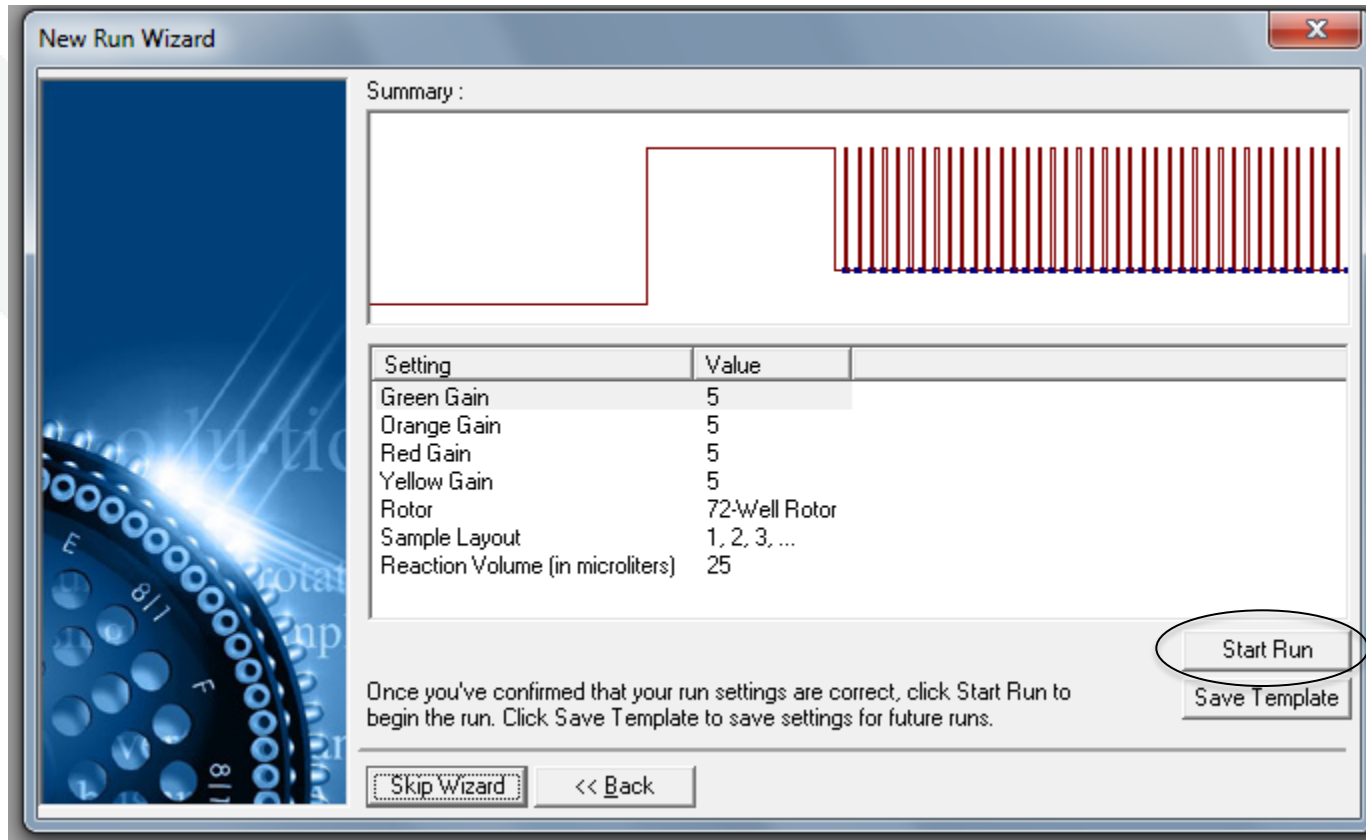
1. Programme the FTD protocol (here: fast-track mastermix)
2. You can add the different steps/ temperatures/ times.

How to edit the set up of a plate. (dyes)



1. After adding a cycling step you need to choose the channels.
2. Tick the pre-installed Acquiring channels Green(Fam), Yellow(Hex-Vic-Yak) Orange(Rox) and Red(Cy5-Atto). Press OK.

How to insert and start a plate.



1. After pressing OK, this window opens.
2. Open the cyclor by shifting the lid backwards and insert the rotor.
3. After inserting your rotor containing your tubes close the lid by sliding forward.
4. Choose Start Run and add the destination of your run file.

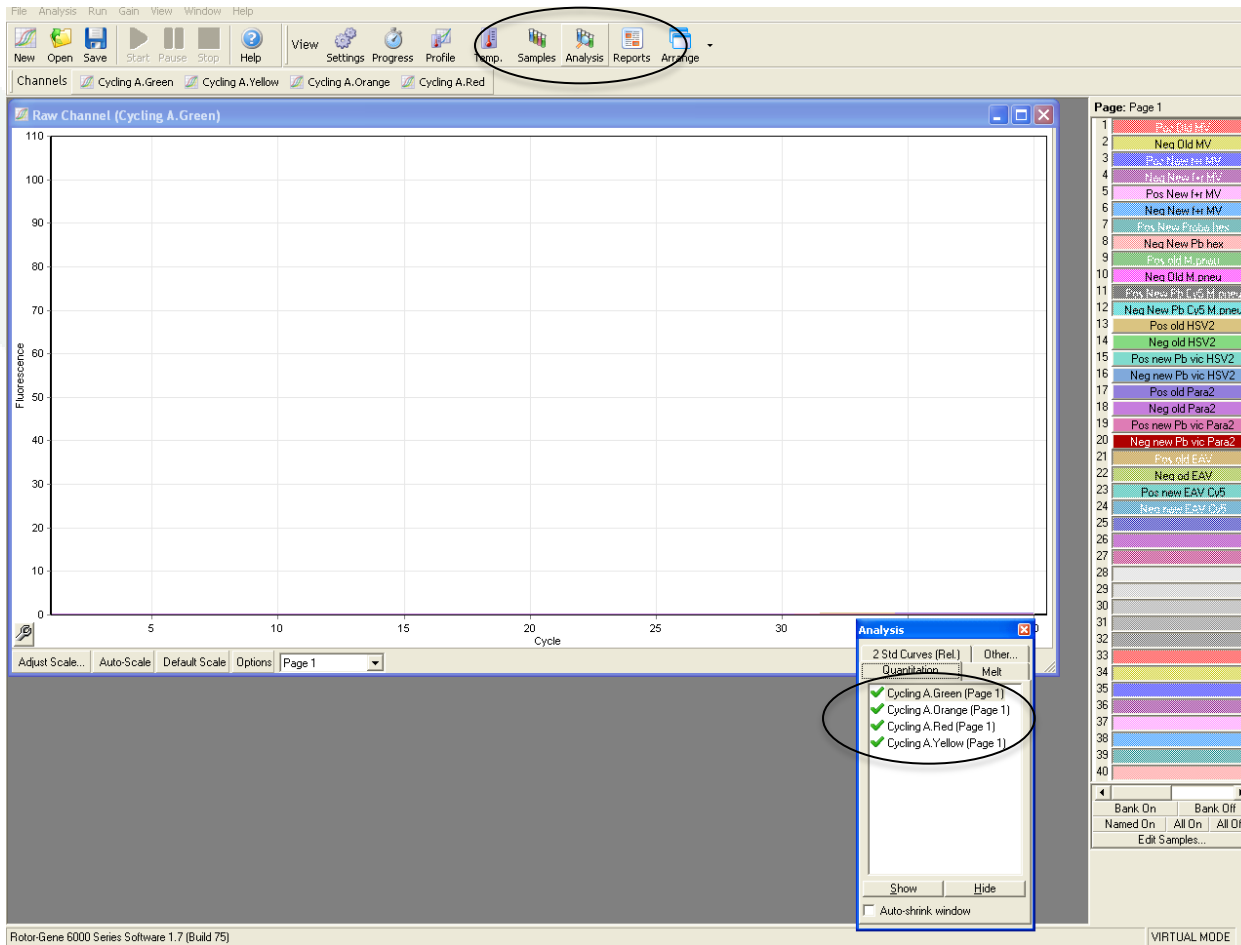
How to edit the set up of a plate. (samples)

The screenshot shows the RotorGene 6000 Series Software 1.7 (Build 75) interface. The main window displays the 'Edit Samples' dialog box, which is used to configure the samples for a run. The dialog box has a menu bar (File, Edit, Format, Security) and a toolbar with icons for New, Open, Save, Start, Pause, Stop, and Help. The 'Settings' section shows 'Standard' selected and 'Rotor Style' chosen. The 'Given Conc. Format' is set to '123.457' and the 'Unit' is 'copies/ul'. The 'Samples' section contains a table with columns for C, ID, Name, Type, Groups, Given Conc., and Selected. The table lists 20 samples, including various controls and positive samples. The 'Page' section at the bottom of the dialog box shows 'Page 1' and buttons for 'Undo', 'OK', 'Cancel', and 'Help'. The 'OK' button is circled in red. To the right of the dialog box, a vertical list of 40 channels is visible, with the first 11 channels corresponding to the samples in the dialog box. The 'Edit Samples...' button in the bottom right corner of the main window is also circled in red.

C	ID	Name	Type	Groups	Given Conc.	Selected
1		Flu/Rhino Alic	Negative Control			No
2		Flu/Rhino PC	Positive Control			No
3		Cor Alic	Negative Control			No
4		Cor Pos	Positive Control			No
5		EAV Alic	Negative Control			No
6		EAV Pos	Positive Control			No
7		BoMpPI Alic	Negative Control			No
8		BoMpPI Pos	Positive Control			No
9		RtEPA Alic	Negative Control			No
10		RtEPA Pos	Positive Control			No
11		Flu Alic	Negative Control			Yes
12		Flu Pos	Positive Control			Yes
13			None			No
14			None			No
15			None			No
16			None			No
17			None			No
18			None			No
19			None			No
20			None			No

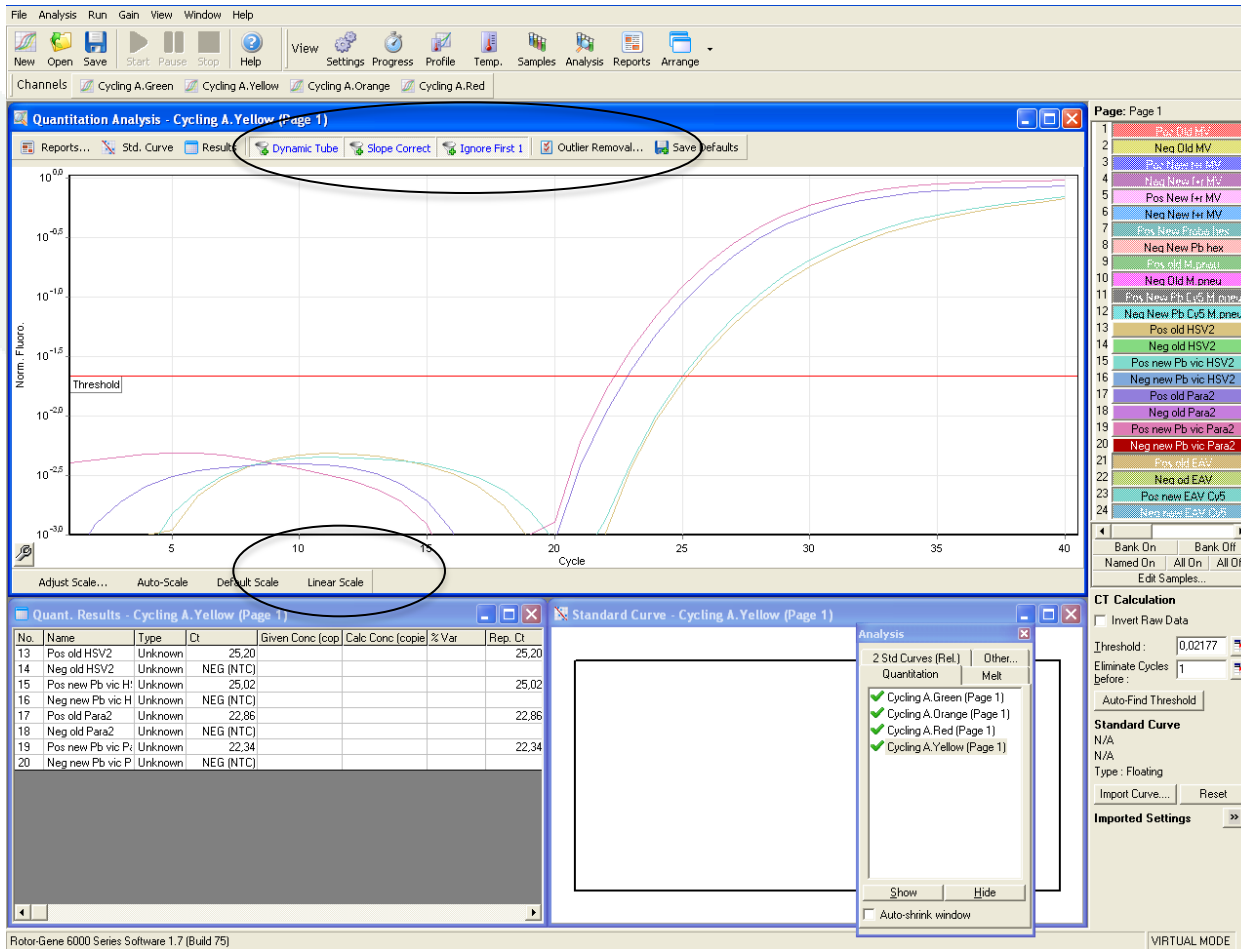
1. After starting your run you choose Edit Sample.
2. Insert your Sample Name and Type and confirm with OK.

How to start analysing your plate.



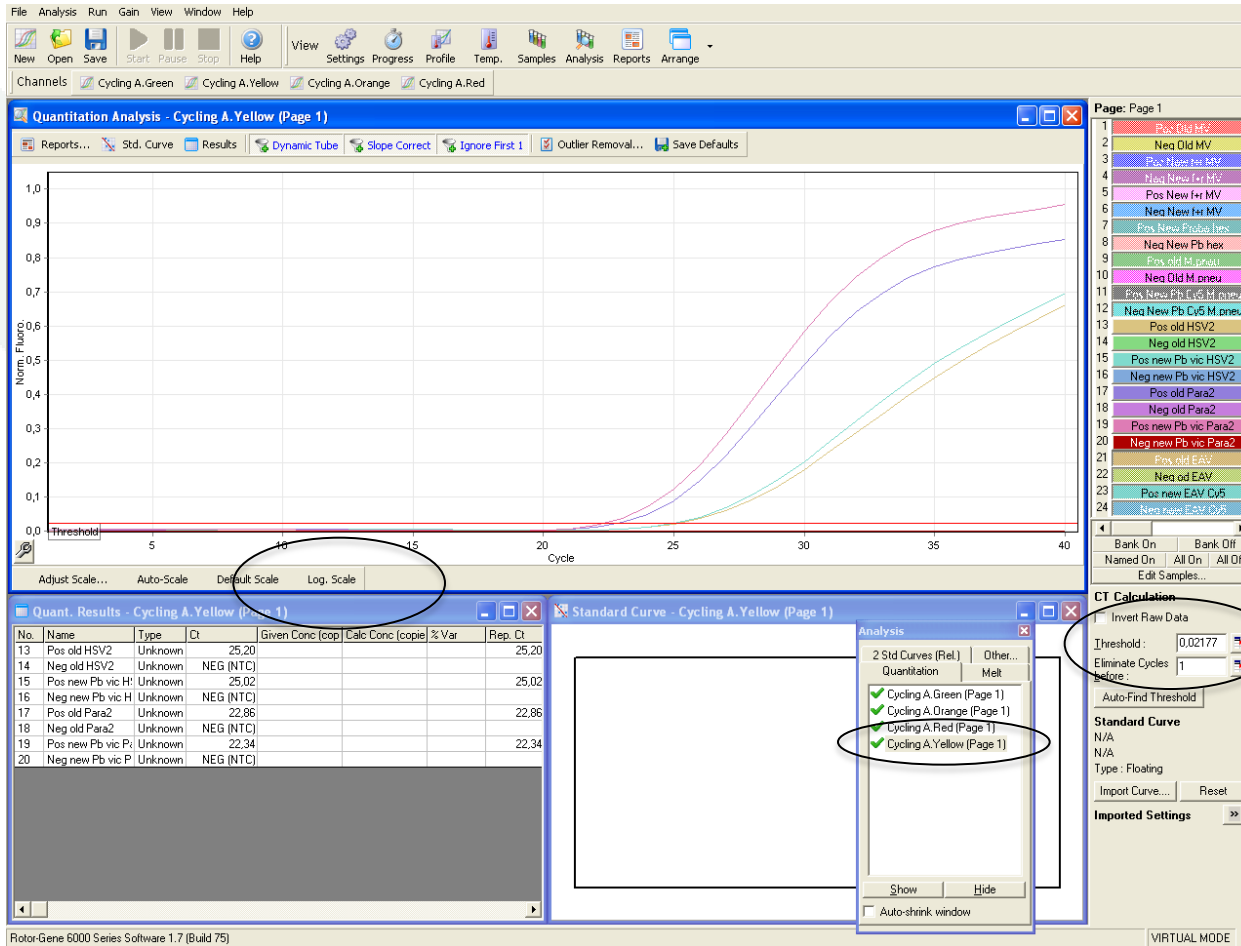
1. After the run has finished you get the raw channel data first.
2. For the RotorGene you need to analyse channel by channel.
3. Tick the Analysis button in the menu above and tick one channel in the box below.

How to start analysing your plate.



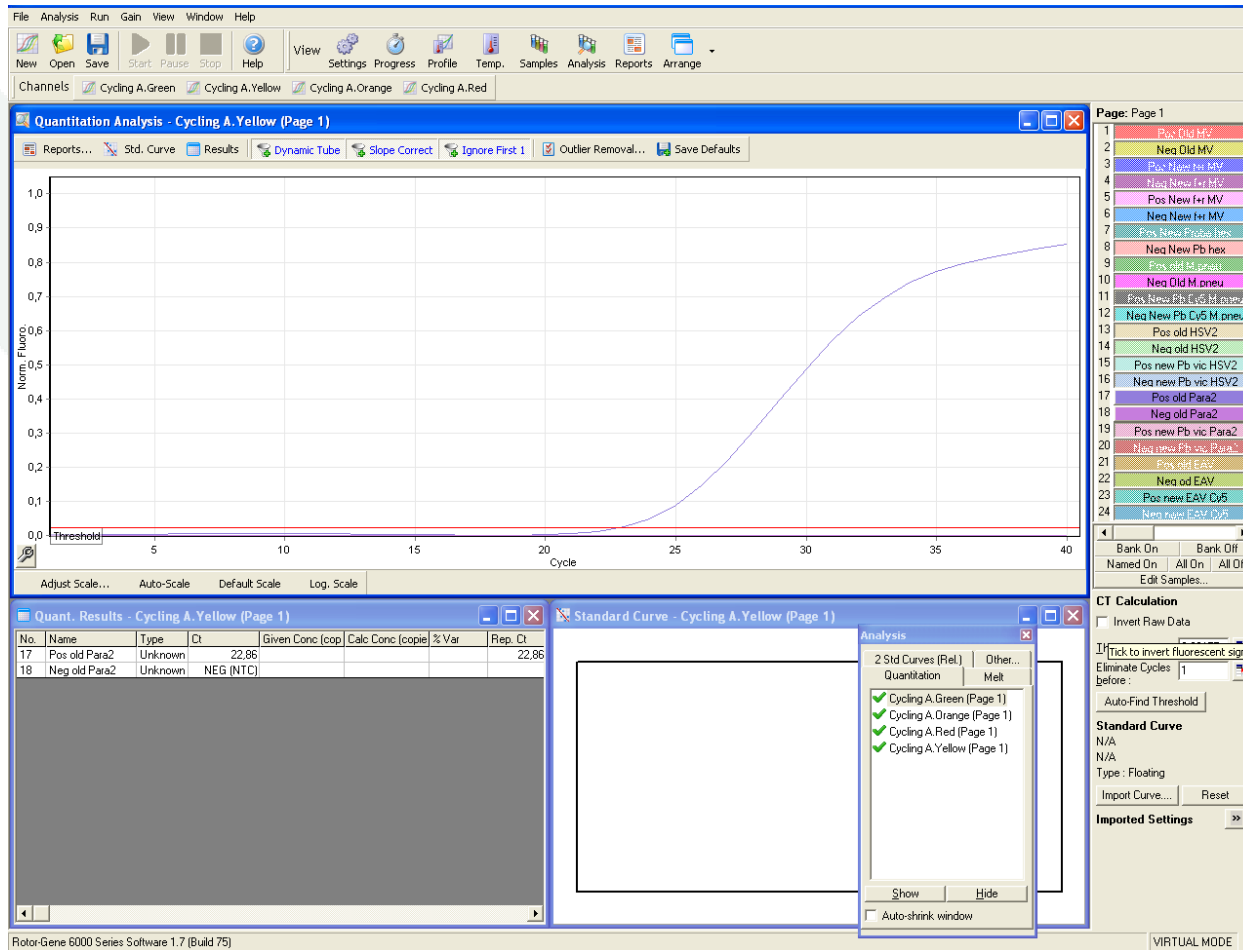
1. Choose Linear Scale and modify the baseline with Slope Correct and Ignore first (e.g. 5). Using Outlier Removal is not recommended, as you could delete low positives.

How to start analysing your plate.



1. In the picture above the Yellow channel was chosen.
2. To modify the threshold you need to click next to the threshold value.
3. In the table view you will find the Ct values.
4. You need to repeat these steps for all 4 channels.

FIRST: Check PC and NC. SECOND: Check patients



1. Now you are able to start with analysing your plate
2. Start with checking you positive, negative and internal control.
3. If your controls are valid (Assay validation-insert notice) you can start to analyse your patient samples.

If you have any further questions or suggestions, please contact us at info@fast-trackdiagnostics.com or check out the FAQ section on our website

www.fast-trackdiagnostics.com